

# Package ‘reportRmd’

November 16, 2023

**Title** Tidy Presentation of Clinical Reporting

**Version** 0.1.0

**Date** 2023-11-10

**Description** Streamlined statistical reporting in 'Rmarkdown' environments.  
Facilitates the automated reporting of descriptive statistics, multiple univariate models, multivariable models and tables combining these outputs. Plotting functions include customisable survival curves, forest plots from logistic and ordinal regression and bivariate comparison plots.

**License** MIT + file LICENSE

**Suggests** rmarkdown, testthat (>= 3.0.0)

**Config/testthat/edition** 3

**Encoding** UTF-8

**RoxygenNote** 7.2.3

**Imports** aod, cmprsk, cowplot, geepack, ggplot2, ggpubr, gridExtra, kableExtra, knitr, lifecycle, MASS, pander, plyr, rlang, rstatix, scales, survival

**Collate** 'helper.R' 'main.R' 'globals.R' 'data.R' 'lblCode.R'

**Depends** R (>= 4.2)

**LazyData** true

**VignetteBuilder** knitr, rmarkdown

**NeedsCompilation** no

**Author** Lisa Avery [cre, aut] (<<https://orcid.org/0000-0002-8431-5143>>),  
Ryan Del Bel [aut],  
Osvaldo Espin-Garcia [aut],  
Katherine Lajkosz [aut] (<<https://orcid.org/0000-0003-3760-5401>>),  
Tyler Pittman [aut] (<<https://orcid.org/0000-0002-5013-6980>>),  
Anna Santiago [aut] (<<https://orcid.org/0000-0002-0932-2386>>),  
Yanning Wang [ctr],  
Jessica Weiss [aut],  
Wei Xu [aut]

**Maintainer** Lisa Avery <lisa.avery@uhn.ca>

Repository CRAN

Date/Publication 2023-11-16 17:00:03 UTC

## R topics documented:

addspace . . . . .	3
boxcofitRx . . . . .	3
cap . . . . .	4
clear_labels . . . . .	4
covsum . . . . .	5
crrRx . . . . .	7
ctDNA . . . . .	8
excelCol . . . . .	8
excelColLetters . . . . .	9
extract_labels . . . . .	10
forestplot2 . . . . .	10
forestplotMV . . . . .	11
forestplotUV . . . . .	13
forestplotUVMV . . . . .	14
formatp . . . . .	16
geoR_boxcofit . . . . .	16
ggkmcif . . . . .	17
ggkmcif2 . . . . .	21
ggkmcif2Parameters . . . . .	23
ggkmcif_paste . . . . .	26
hbld . . . . .	27
lbld . . . . .	27
lpvalue . . . . .	27
mvsum . . . . .	28
nestTable . . . . .	29
nicename . . . . .	30
niceNum . . . . .	31
outTable . . . . .	31
pembrolizumab . . . . .	33
plotuv . . . . .	34
psthr . . . . .	36
pstprn . . . . .	36
pvalue . . . . .	36
rmds . . . . .	37
rm_cifsum . . . . .	37
rm_covsum . . . . .	39
rm_mvsum . . . . .	42
rm_survdiff . . . . .	44
rm_survsum . . . . .	45
rm_survtime . . . . .	47
rm_uvsum . . . . .	49
rm_uv_mv . . . . .	52

<code>addspace</code>	3
<code>sanitizestr</code> . . . . .	54
<code>set_labels</code> . . . . .	54
<code>set_var_labels</code> . . . . .	55
<code>testData</code> . . . . .	56
<code>uvsum</code> . . . . .	57
<b>Index</b>	<b>59</b>

---

<code>addspace</code>	<i>Add spaces to strings in LaTeX</i>
-----------------------	---------------------------------------

---

**Description**

Add spaces to strings in LaTeX. Returns appends ~~~ before the string

**Usage**

`addspace(x)`

**Arguments**

<code>x</code>	string
----------------	--------

---

<code>boxcoxfitRx</code>	<i>fit box cox transformed linear model</i>
--------------------------	---

---

**Description**

Wrapper function to fit fine and gray competing risk model using function `crr` from package `cmprsk`

**Usage**

`boxcoxfitRx(f, data, lambda = FALSE)`

**Arguments**

<code>f</code>	formula for the model. Currently the formula only works by using the name of the column in a dataframe. It does not work by using \$ or [] notation.
<code>data</code>	dataframe containing data
<code>lambda</code>	boolean indicating if you want to output the lamda used in the boxcox transformation. If so the function will return a list of length 2 with the model as the first element and a vector of length 2 as the second.

**Value**

a list containing the linear model (`lm`) object and, if requested, `lambda`

---

cap	<i>Capitalize a string</i>
-----	----------------------------

---

**Description**

Capitalize a string

**Usage**

```
cap(x)
```

**Arguments**

x	string
---	--------

---

clear_labels	<i>Clear variable labels from a data frame</i>
--------------	--

---

**Description**

This function will remove all label attributes from variables in the data.

**Usage**

```
clear_labels(data)
```

**Arguments**

data	the data frame to remove labels from
------	--------------------------------------

**Details**

To change or remove individual labels use `set_labels` or `set_var_labels`

**Examples**

```
# Set a few variable labels for ctDNA
ctDNA <- ctDNA |> set_var_labels(
  ctdna_status="detectable ctDNA",
  cohort="A cohort label")
# Clear all variable data frames and check
clear_labels(ctDNA)
```

---

covsum	<i>Get covariate summary dataframe</i>
--------	--

---

### Description

Returns a dataframe corresponding to a descriptive table.

### Usage

```
covsum(
  data,
  covs,
  maincov = NULL,
  digits = 1,
  numobs = NULL,
  markup = TRUE,
  sanitize = TRUE,
  nicenames = TRUE,
  IQR = FALSE,
  all.stats = FALSE,
  pvalue = TRUE,
  effSize = FALSE,
  show.tests = FALSE,
  dropLevels = TRUE,
  excludeLevels = NULL,
  full = TRUE,
  digits.cat = 0,
  testcont = c("rank-sum test", "ANOVA"),
  testcat = c("Chi-squared", "Fisher"),
  include_missing = FALSE,
  percentage = c("column", "row")
)
```

### Arguments

<code>data</code>	dataframe containing data
<code>covs</code>	character vector with the names of columns to include in table
<code>maincov</code>	covariate to stratify table by
<code>digits</code>	number of digits for summarizing mean data, does not affect p-values
<code>numobs</code>	named list overriding the number of people you expect to have the covariate
<code>markup</code>	boolean indicating if you want latex markup
<code>sanitize</code>	boolean indicating if you want to sanitize all strings to not break LaTeX
<code>nicenames</code>	boolean indicating if you want to replace . and _ in strings with a space
<code>IQR</code>	boolean indicating if you want to display the inter quantile range (Q1,Q3) as opposed to (min,max) in the summary for continuous variables

<code>all.stats</code>	boolean indicating if all summary statistics (Q1,Q3 + min,max on a separate line) should be displayed. Overrides IQR.
<code>pvalue</code>	boolean indicating if you want p-values included in the table
<code>effSize</code>	boolean indicating if you want effect sizes included in the table. Can only be obtained if <code>pvalue</code> is also requested. Effect sizes calculated include Cramer's V for categorical variables, Cohen's d, Wilcoxon r, or Eta-squared for numeric/continuous variables.
<code>show.tests</code>	boolean indicating if the type of statistical test and effect size used should be shown in a column beside the pvalues. Ignored if <code>pvalue=FALSE</code> .
<code>dropLevels</code>	logical, indicating if empty factor levels be dropped from the output, default is TRUE.
<code>excludeLevels</code>	a named list of covariate levels to exclude from statistical tests in the form <code>list(varname =c('level1','level2'))</code> . These levels will be excluded from association tests, but not the table. This can be useful for levels where there is a logical skip (ie not missing, but not presented). Ignored if <code>pvalue=FALSE</code> .
<code>full</code>	boolean indicating if you want the full sample included in the table, ignored if <code>maincov</code> is NULL
<code>digits.cat</code>	number of digits for the proportions when summarizing categorical data (default: 0)
<code>testcont</code>	test of choice for continuous variables,one of <i>rank-sum</i> (default) or <i>ANOVA</i>
<code>testcat</code>	test of choice for categorical variables,one of <i>Chi-squared</i> (default) or <i>Fisher</i>
<code>include_missing</code>	Option to include NA values of <code>maincov</code> . NAs will not be included in statistical tests
<code>percentage</code>	choice of how percentages are presented ,one of <i>column</i> (default) or <i>row</i>

## Details

Comparisons for categorical variables default to chi-square tests, but if there are counts of <5 then the Fisher Exact test will be used and if this is unsuccessful then a second attempt will be made computing p-values using MC simulation. If `testcont='ANOVA'` then the t-test with unequal variance will be used for two groups and an ANOVA will be used for three or more. The statistical test used can be displayed by specifying `show.tests=TRUE`.

The number of decimals places to display the statistics can be changed with `digits`, but this will not change the display of p-values. If more significant digits are required for p-values then use `tableOnly=TRUE` and format as desired.

## References

- Ellis, P.D. (2010) The essential guide to effect sizes: statistical power, meta-analysis, and the interpretation of research results. Cambridge: Cambridge University Press.[doi:10.1017/CBO9780511761676](https://doi.org/10.1017/CBO9780511761676)
- Lakens, D. (2013) Calculating and reporting effect sizes to facilitate cumulative science: a practical primer for t-tests and ANOVAs. *Frontiers in Psychology*, 4; 863:1-12. [doi:10.3389/fpsyg.2013.00863](https://doi.org/10.3389/fpsyg.2013.00863)

**See Also**

[fisher.test](#), [chisq.test](#), [wilcox.test](#), [kruskal.test](#), and [anova](#)

---

crrRx

*fit crr model*

---

**Description**

Wrapper function to fit fine and gray competing risk model using function `crr` from package `cmprsk`

**Usage**

```
crrRx(f, data)
```

**Arguments**

<code>f</code>	formula for the model. Currently the formula only works by using the name of the column in a dataframe. It does not work by using <code>\$</code> or <code>[]</code> notation.
<code>data</code>	dataframe containing data

**Value**

a competing risk model with the call appended to the list

**See Also**

[crr](#)

**Examples**

```
# From the crr help file:
set.seed(10)
ftime <- rexp(200)
fstatus <- sample(0:2,200,replace=TRUE)
cov <- matrix(runif(600),nrow=200)
dimnames(cov)[[2]] <- c('x1','x2','x3')
df <- data.frame(ftime,fstatus,cov)
m1 <- crrRx(as.formula('ftime+fstatus~x1+x2+x3'),df)
# Nicely output to report:
rm_mvsum(m1,data=df,showN = TRUE,vif=TRUE)
```

---

ctDNA	<i>Tumour size change over time Longitudinal changes in tumour size since baseline for patients by changes in ctDNA status (clearance, decrease or increase) since baseline.</i>
-------	--

---

**Description**

Tumour size change over time

Longitudinal changes in tumour size since baseline for patients by changes in ctDNA status (clearance, decrease or increase) since baseline.

**Usage**

ctDNA

**Format**

A data frame with 270 rows and 5 variables:

**id** Patient ID

**cohort** Study Cohort: A = Squamous cell carcinoma of soft pallate, B = Triple negative breast cancer, C = Ovarian, high grade serous, D = Melanoma, E = Other Solid Tumor

**ctdna\_status** Change in ctDNA since baseline

**time** Number of weeks on treatment

**size\_change** Percentage change in tumour measurement

**Source**

<https://www.nature.com/articles/s43018-020-0096-5>

---

excelCol	<i>Retrieve columns number from spreadsheet columns specified as unquoted letters</i>
----------	---

---

**Description**

Retrieve columns number from spreadsheet columns specified as unquoted letters

**Usage**

excelCol(...)

**Arguments**

... unquoted excel column headers (i.e. excelCol(A,CG,AA)) separated by commas



**Value**

a numeric vector corresponding to columns in a spreadsheet

**Examples**

```
## Find the column numbers for excel columns AB, CE and BB
excelCol(AB,CE,bb)
## Get the columns between A and K and Z
excelCol(A-K,Z)
```

---

excelColLetters	<i>Retrieve spreadsheet column letter-names from columns indices</i>
-----------------	--

---

**Description**

Creates a vector of spreadsheet-style letter-names corresponding to column numbers

**Usage**

```
excelColLetters(columnIndices)
```

**Arguments**

columnIndices    vector of integer column indices

**Details**

This is the inverse function of excelCol

**Value**

a character vector corresponding to the spreadsheet column headings

**Examples**

```
## Find the column numbers for excel columns AB, CE and BB
colIndices <- excelCol(AB,CE,bb)
## Go back to the column names
excelColLetters(colIndices)
```

---

extract_labels	<i>Extract variable labels from labelled data frame</i>
----------------	---

---

**Description**

Extract variable labels from data and return a data frame with labels

**Usage**

```
extract_labels(data, sep = "_")
```

**Arguments**

data	the data frame to extract labels from
sep	character used to separate multiple labels, defaults to "_"

**Details**

All variable names will be returned, even those with no labels. If the label attribute has length greater than one the values will be concatenated and returned as a single string separated by sep

**Examples**

```
# Set a few variable labels for ctDNA
ctDNA <- ctDNA |> set_var_labels(
  ctdna_status="detectable ctDNA",
  cohort="A cohort label")
# Extract labels
extract_labels(ctDNA)
```

---

forestplot2	<i>Create a forest plot using ggplot2</i>
-------------	---

---

**Description**

This function will accept a log or logistic regression fit from glm or geeglm, and display the OR or RR for each variable on the appropriate log scale.

**Usage**

```
forestplot2(
  model,
  conf.level = 0.95,
  orderByRisk = TRUE,
  colours = "default",
  showEst = TRUE,
```

```

    rmRef = FALSE,
    logScale = getOption("reportRmd.logScale", TRUE),
    nxTicks = 5
  )

```

### Arguments

model	an object output from the glm or geeglm function, must be from a logistic regression
conf.level	controls the width of the confidence interval
orderByRisk	logical, should the plot be ordered by risk
colours	can specify colours for risks less than, 1 and greater than 1.0. Default is red, black, green
showEst	logical, should the risks be displayed on the plot in text
rmRef	logical, should the reference levels be removed for the plot?
logScale	logical, should OR/RR be shown on log scale, defaults to TRUE, or reportRmd.logScale if set. See <a href="https://doi.org/10.1093/aje/kwr156">https://doi.org/10.1093/aje/kwr156</a> for why you may prefer a linear scale.
nxTicks	Number of tick marks supplied to the log_breaks function to produce

### Value

a plot object

### Examples

```

data("pembrolizumab")
glm_fit = glm(orr~change_ctdna_group+sex+age+l_size,
data=pembrolizumab,family = 'binomial')
forestplot2(glm_fit)

```

---

forestplotMV

---

*Create a multivariable forest plot using ggplot2*


---

### Description

This function will send and take log or logistic regression fit from glm or geeglm from mvsum function, and display the OR or RR for each variable on the appropriate log scale.

**Usage**

```
forestplotMV(
  model,
  data,
  conf.level = 0.95,
  orderByRisk = TRUE,
  colours = "default",
  showEst = TRUE,
  rmRef = FALSE,
  digits = getOption("reportRmd.digits", 2),
  logScale = getOption("reportRmd.logScale", TRUE),
  nxTicks = 5,
  showN = TRUE,
  showEvent = TRUE
)
```

**Arguments**

model	an object output from the glm or geeglm function, must be from a logistic regression
data	dataframe containing your data
conf.level	controls the width of the confidence interval
orderByRisk	logical, should the plot be ordered by risk
colours	can specify colours for risks less than, 1 and greater than 1.0. Default is red, black, green
showEst	logical, should the risks be displayed on the plot in text
rmRef	logical, should the reference levels be removed for the plot?
digits	number of digits to use displaying estimates
logScale	logical, should OR/RR be shown on log scale, defaults to TRUE, or reportRmd.logScale if set. See <a href="https://doi.org/10.1093/aje/kwr156">https://doi.org/10.1093/aje/kwr156</a> for why you may prefer a linear scale.
nxTicks	Number of tick marks supplied to the log_breaks function to produce
showN	Show number of observations per variable and category
showEvent	Show number of events per variable and category

**Value**

a plot object

**Examples**

```
data("pembrolizumab")
glm_fit = glm(orr~change_ctdna_group+sex+age+l_size,
  data=pembrolizumab,family = 'binomial')
forestplotMV(glm_fit)
```

---

forestplotUV

*Create an univariable forest plot using ggplot2*


---

### Description

This function will send and take log or logistic regression fit from glm or geeglm from uvsum function, and display the OR or RR for each variable on the appropriate log scale.

### Usage

```
forestplotUV(
  response,
  covs,
  data,
  id = NULL,
  corstr = NULL,
  model = "glm",
  family = NULL,
  digits = getOption("reportRmd.digits", 2),
  conf.level = 0.95,
  orderByRisk = TRUE,
  colours = "default",
  showEst = TRUE,
  rmRef = FALSE,
  logScale = getOption("reportRmd.logScale", TRUE),
  nxTicks = 5,
  showN = TRUE,
  showEvent = TRUE
)
```

### Arguments

response	character vector with names of columns to use for response
covs	character vector with names of columns to use for covariates
data	dataframe containing your data
id	character vector which identifies clusters. Only used for geeglm
corstr	character string specifying the correlation structure. Only used for geeglm. The following are permitted: "independence", "exchangeable", "ar1", "unstructured" and "userdefined"
model	fitted model object
family	description of the error distribution and link function to be used in the model. Only used for geeglm
digits	number of digits to round to
conf.level	controls the width of the confidence interval

orderByRisk	logical, should the plot be ordered by risk
colours	can specify colours for risks less than, 1 and greater than 1.0. Default is red, black, green
showEst	logical, should the risks be displayed on the plot in text
rmRef	logical, should the reference levels be removed for the plot?
logScale	logical, should OR/RR be shown on log scale, defaults to TRUE, or reportRmd.logScale if set. See <a href="https://doi.org/10.1093/aje/kwr156">https://doi.org/10.1093/aje/kwr156</a> for why you may prefer a linear scale.
nxTicks	Number of tick marks supplied to the log_breaks function to produce
showN	Show number of observations per variable and category
showEvent	Show number of events per variable and category

**Value**

a plot object

**Examples**

```
data("pembrolizumab")
forestplotUV(response="orr", covs=c("change_ctdna_group", "sex", "age", "l_size"),
data=pembrolizumab, family='binomial')
```

---

forestplotUVMV

---

*Combine an univariable and multivariable forest plot using ggplot2*


---

**Description**

This function will take log or logistic regression fit forest plot output from forestplotUV and forestplotMV functions and display the combined adjusted and unadjusted OR or RR for each variable on the appropriate log scale. Please note that total N and reference-level N is taken from unadjusted model.

**Usage**

```
forestplotUVMV(
  UVmodel,
  MVmodel,
  model = "glm",
  family = NULL,
  digits = getOption("reportRmd.digits", 2),
  orderByRisk = TRUE,
  colours = "default",
  showEst = TRUE,
  rmRef = FALSE,
  logScale = FALSE,
```

```

    nxTicks = 5,
    showN = TRUE,
    showEvent = TRUE
  )

```

### Arguments

UVmodel	an UV model object output from the forestplotUV function
MVmodel	a MV model object output from the forestplotMV function
model	fitted model object
family	description of the error distribution and link function to be used in the model. Only used for geeglm
digits	number of digits to round to
orderByRisk	logical, should the plot be ordered by risk
colours	can specify colours for risks less than, 1 and greater than 1.0. Default is red, black, green
showEst	logical, should the risks be displayed on the plot in text
rmRef	logical, should the reference levels be removed for the plot?
logScale	logical, should OR/RR be shown on log scale, defaults to TRUE. See <a href="https://doi.org/10.1093/aje/kwr156">https://doi.org/10.1093/aje/kwr156</a> for why you may prefer a linear scale.
nxTicks	Number of tick marks supplied to the log_breaks function to produce
showN	Show number of observations per variable and category
showEvent	Show number of events per variable and category

### Value

a plot object

### Examples

```

data("pembrolizumab")
UVp = forestplotUV(response="orr", covs=c("change_ctdna_group", "sex", "age",
    "l_size"), data=pembrolizumab, family='binomial')
MVp = forestplotMV(glm(orr~change_ctdna_group+sex+age+l_size,
    data=pembrolizumab,family = 'binomial'))
forestplotUVMV(UVp, MVp)

```

---

formatp	<i>Specific p-value formatting</i>
---------	------------------------------------

---

**Description**

If  $p < 0.001$  returns "<0.001", if  $p < 0.01$  returns p to 3 decimal places otherwise returns p to 2 decimal places

**Usage**

```
formatp(pvalues)
```

**Arguments**

pvalues	a vector of p values
---------	----------------------

---

geoR_boxcoffit	<i>Parameter Estimation for the Box-Cox Transformation</i>
----------------	--

---

**Description**

This function is copied from the geoR package which has been removed from the CRAN repository.

**Usage**

```
geoR_boxcoffit(object, xmat, lambda, lambda2 = NULL, add.to.data = 0)
```

**Arguments**

object	a vector with the data
xmat	a matrix with covariates values. Defaults to rep(1, length(y)).
lambda	numerical value(s) for the transformation parameter lambda. Used as the initial value in the function for parameter estimation. If not provided default values are assumed. If multiple values are passed the one with highest likelihood is used as initial value.
lambda2	logical or numerical value(s) of the additional transformation (see DETAILS below). Defaults to NULL. If TRUE this parameter is also estimated and the initial value is set to the absolute value of the minimum data. A numerical value is provided it is used as the initial value. Multiple values are allowed as for lambda.
add.to.data	a constant value to be added to the data.

**Details**

For more information see: <https://cran.r-project.org/web/packages/geoR/index.html>



---

`ggkmcif`*Plot KM and CIF curves with ggplot*

---

**Description**

This function will plot a KM or CIF curve with option to add the number at risk. You can specify if you want confidence bands, the hazard ratio, and pvalues, as well as the units of time used.

**Usage**

```
ggkmcif(  
  response,  
  cov = NULL,  
  data,  
  type = NULL,  
  pval = TRUE,  
  HR = FALSE,  
  HR_pval = FALSE,  
  conf.curves = FALSE,  
  conf.type = "log",  
  table = TRUE,  
  times = NULL,  
  xlab = "Time",  
  ylab = NULL,  
  main = NULL,  
  stratalabs = NULL,  
  strataname = nicename(cov),  
  stratalabs.table = NULL,  
  strataname.table = strataname,  
  median.text = FALSE,  
  median.lines = FALSE,  
  median.CI = FALSE,  
  set.time.text = NULL,  
  set.time.line = FALSE,  
  set.time = 5,  
  set.time.CI = FALSE,  
  censor.marks = TRUE,  
  censor.size = 3,  
  censor.stroke = 1.5,  
  fsize = 10,  
  nsize = 3,  
  lsize = 1,  
  psize = 3.5,  
  median.size = 3,  
  median.pos = NULL,  
  median.lsize = 1,  
  set.size = 3,  
)
```

```

set.pos = NULL,
set.lsize = 1,
ylim = c(0, 1),
col = NULL,
linetype = NULL,
xlim = NULL,
legend.pos = NULL,
pval.pos = NULL,
plot.event = 1,
event = c("col", "linetype"),
flip.CIF = FALSE,
cut = NULL,
eventlabs = NULL,
event.name = NULL,
Numbers_at_risk_text = "Numbers at risk",
HR.digits = 2,
HR.pval.digits = 3,
pval.digits = 3,
median.digits = 3,
set.time.digits = 3,
returns = FALSE,
print.n.missing = TRUE
)

```

### Arguments

response	character vector with names of columns to use for response
cov	String specifying the column name of stratification variable
data	dataframe containing your data
type	string indicating the type of univariate model to fit. The function will try and guess what type you want based on your response. If you want to override this you can manually specify the type. Options include "KM", and "CIF"
pval	boolean to specify if you want p-values in the plot (Log Rank test for KM and Gray's test for CIF)
HR	boolean to specify if you want hazard ratios included in the plot
HR_pval	boolean to specify if you want HR p-values in the plot
conf.curves	boolean to specify if you want confidence interval bands
conf.type	One of "none"(the default), "plain", "log", "log-log" or "logit". Only enough of the string to uniquely identify it is necessary. The first option causes confidence intervals not to be generated. The second causes the standard intervals curve $\pm k * se(\text{curve})$ , where k is determined from conf.int. The log option calculates intervals based on the cumulative hazard or log(survival). The log-log option bases the intervals on the log hazard or log(-log(survival)), and the logit option on $\log(\text{survival}/(1-\text{survival}))$ .
table	Logical value. If TRUE, includes the number at risk table
times	Numeric vector of times for the x-axis

xlab	String corresponding to xlabel. By default is "Time"
ylab	String corresponding to ylabel. When NULL uses "Survival probability" for KM curves, and "Probability of an event" for CIF
main	String corresponding to main title. When NULL uses Kaplan-Meier Plots, and "Cumulative Incidence Plot for CIF"
stratalabs	string corresponding to the labels of the covariate, when NULL will use the levels of the covariate
strataname	String of the covariate name default is nicename(cov)
stratalabs.table	String corresponding to the levels of the covariate for the number at risk table, when NULL will use the levels of the covariate. Can use a string of "-" when the labels are long
strataname.table	String of the covariate name for the number at risk table default is nicename(cov)
median.text	boolean to specify if you want the median values added to the legend (or as added text if there are no covariates), for KM only
median.lines	boolean to specify if you want the median values added as lines to the plot, for KM only
median.CI	boolean to specify if you want the 95% with the median text (Only for KM)
set.time.text	string for the text to add survival at a specified time (eg. year OS)
set.time.line	boolean to specify if you want the survival added as lines to the plot at a specified point
set.time	Numeric values of the specific time of interest, default is 5 (Multiple values can be entered)
set.time.CI	boolean to specify if you want the 95% interval with the set time text
censor.marks	logical value. If TRUE, includes censor marks (only for KM curves)
censor.size	size of censor marks, default is 3
censor.stroke	stroke of censor marks, default is 1.5
fsize	font size
nsize	font size for numbers in the numbers at risk table
lsize	line size
psize	size of the pvalue
median.size	size of the median text (Only when there are no covariates)
median.pos	vector of length 2 corresponding to the median position (Only when there are no covariates)
median.lsize	line size of the median lines
set.size	size of the survival at a set time text (Only when there are no covariates)
set.pos	vector of length 2 corresponding to the survival at a set point position (Only when there are no covariates)
set.lsize	line size of the survival at set points

<code>ylim</code>	vector of length 2 corresponding to limits of y-axis. Default to NULL
<code>col</code>	vector of colours
<code>linetype</code>	vector of line types
<code>xlim</code>	vector of length 2 corresponding to limits of x-axis. Default to NULL
<code>legend.pos</code>	Can be either a string corresponding to the legend position ("left", "top", "right", "bottom", "none") or a vector of length 2 corresponding to the legend position (uses normalized units (ie the <code>c(0.5,0.5)</code> is the middle of the plot))
<code>pval.pos</code>	vector of length 2 corresponding to the p-value position
<code>plot.event</code>	Which event(s) to plot (1,2, or <code>c(1,2)</code> )
<code>event</code>	String specifying if the event should be mapped to the colour, or linetype when plotting both events to colour = "col", line type
<code>flip.CIF</code>	boolean to flip the CIF curve to start at 1
<code>cut</code>	numeric value indicating where to divide a continuous covariate (default is the median)
<code>eventlabs</code>	String corresponding to the event type names
<code>event.name</code>	String corresponding to the label of the event types
<code>Numbers_at_risk_text</code>	String for the label of the number at risk
<code>HR.digits</code>	Number of digits printed of the hazard ratio
<code>HR.pval.digits</code>	Number of digits printed of the hazard ratio pvalue
<code>pval.digits</code>	Number of digits printed of the Gray's/log rank pvalue
<code>median.digits</code>	Number of digits printed of the median pvalue
<code>set.time.digits</code>	Number of digits printed of the probability at a specified time
<code>returns</code>	Logical value returns a list with all ggplot objects in a list
<code>print.n.missing</code>	Logical, should the number of missing be shown !Needs to be checked

## Details

Note that for proper pdf output of special characters the following code needs to be included in the first chunk of the rmd `knitr::opts_chunk$set(dev="cairo_pdf")`

## Value

Nothing is returned unless `returns = TRUE` is used. With `returns = TRUE`, if `table=TRUE` (the default) a table style graphic with survival plot and number at risk table is returned. Otherwise a plot with the survival curves is returned.

**Examples**

```

data("pembrolizumab")
# Simple plot without confidence intervals
ggkmcif(response = c('os_time','os_status'),
cov='cohort',
data=pembrolizumab)

# Plot with median survival time
ggkmcif(response = c('os_time','os_status'),
cov='sex',
data=pembrolizumab,
median.text = TRUE,median.lines=TRUE,conf.curves=TRUE)

# Plot with specified survival times and log-log CI
ggkmcif(response = c('os_time','os_status'),
cov='sex',
data=pembrolizumab,
median.text = FALSE,set.time.text = 'mo OS',
set.time = c(12,24),conf.type = 'log-log',conf.curves=TRUE)

# KM plot with 95% CI and censor marks
ggkmcif(c('os_time','os_status'),'sex',data = pembrolizumab, type = 'KM',
HR=TRUE, HR_pval = TRUE, conf.curves = TRUE,conf.type='log-log',
set.time.CI = TRUE, censor.marks=TRUE)

```

ggkmcif2

*Plot KM and CIF curves with ggplot***Description**

This function will plot a KM or CIF curve with option to add the number at risk. You can specify if you want confidence bands, the hazard ratio, and pvalues, as well as the units of time used.

**Usage**

```

ggkmcif2(
  response,
  cov = NULL,
  data,
  pval = TRUE,
  conf.curves = FALSE,
  table = TRUE,
  xlab = "Time",
  ylab = NULL,
  col = NULL,
  times = NULL,
  type = NULL,
  plot.event = 1,
  ...
)

```

**Arguments**

response	character vector with names of columns to use for response
cov	String specifying the column name of stratification variable
data	dataframe containing your data
pval	boolean to specify if you want p-values in the plot (Log Rank test for KM and Gray's test for CIF)
conf.curves	boolean to specify if you want confidence interval bands
table	Logical value. If TRUE, includes the number at risk table
xlab	String corresponding to xlabel. By default is "Time"
ylab	String corresponding to ylabel. When NULL uses "Survival
col	vector of colours
times	Numeric vector of times for the x-axis probability" for KM cuves, and "Probability of an event" for CIF
type	string indicating he type of univariate model to fit. The function will try and guess what type you want based on your response. If you want to override this you can manually specify the type. Options include "KM", and ,"CIF"
plot.event	Which event(s) to plot (1,2, or c(1,2))
...	additional plotting arguments see <a href="#">ggkmCIF2Parameters</a>

**Details**

Note that for proper pdf output of special characters the following code needs to be included in the first chunk of the rmd knitr::opts\_chunk\$set(dev="cairo\_pdf")

**Value**

ggplot object; if table = F then only curves are output; if table = T then curves and risk table are output together

**Examples**

```
# Simple plot without confidence intervals
data("pembrolizumab")
ggkmCIF2(response = c('os_time', 'os_status'),
cov='cohort',
data=pembrolizumab)

# Plot with median survival time
ggkmCIF2(response = c('os_time', 'os_status'),
cov='sex',
data=pembrolizumab,
median.text = TRUE, median.lines=TRUE, conf.curves=TRUE)

# Plot with specified survival times and log-log CI
ggkmCIF2(response = c('os_time', 'os_status'),
cov='sex',
```

```
data=pembrolizumab,
median.text = FALSE,set.time.text = 'mo OS',
set.time = c(12,24),conf.type = 'log-log',conf.curves=TRUE)

# KM plot with 95% CI and censor marks
ggkmCIF2(c('os_time','os_status'),'sex',data = pembrolizumab, type = 'KM',
HR=TRUE, HR_pval = TRUE, conf.curves = TRUE,conf.type='log-log',
set.time.CI = TRUE, censor.marks=TRUE)
```

---

ggkmCIF2Parameters      *Additional parameters passed to ggkmCIF2*

---

## Description

Additional parameters passed to ggkmCIF2

## Usage

```
ggkmCIF2Parameters(
  table.height = NULL,
  HR = FALSE,
  HR_pval = FALSE,
  conf.type = "log",
  main = NULL,
  stratalabs = NULL,
  strataname,
  stratalabs.table = NULL,
  strataname.table = strataname,
  median.text = FALSE,
  median.lines = FALSE,
  median.CI = FALSE,
  set.time.text = NULL,
  set.time.line = FALSE,
  set.time = 5,
  set.time.CI = FALSE,
  censor.marks = TRUE,
  censor.size = 3,
  censor.stroke = 1.5,
  fsize = 11,
  nsize = 3,
  lsize = 1,
  psize = 3.5,
  median.size = 3,
  median.pos = NULL,
  median.lsize = 1,
  set.size = 3,
  set.pos = NULL,
  set.lsize = 1,
```

```

ylim = c(0, 1),
linetype = NULL,
xlim = NULL,
legend.pos = NULL,
pval.pos = NULL,
event = c("col", "linetype"),
flip.CIF = FALSE,
cut = NULL,
eventlabs = NULL,
event.name = NULL,
Numbers_at_risk_text = "Number at risk",
HR.digits = 2,
HR.pval.digits = 3,
pval.digits = 3,
median.digits = 3,
set.time.digits = 3,
print.n.missing = TRUE
)

```

### Arguments

table.height	Relative height of risk table (0-1)
HR	boolean to specify if you want hazard ratios included in the plot
HR_pval	boolean to specify if you want HR p-values in the plot
conf.type	One of "none"(the default), "plain", "log", "log-log" or "logit". Only enough of the string to uniquely identify it is necessary. The first option causes confidence intervals not to be generated. The second causes the standard intervals curve $\pm k * se(\text{curve})$ , where k is determined from conf.int. The log option calculates intervals based on the cumulative hazard or log(survival). The log-log option bases the intervals on the log hazard or log(-log(survival)), and the logit option on log(survival/(1-survival)).
main	String corresponding to main title. When NULL uses Kaplan-Meier Plot s, and "Cumulative Incidence Plot for CIF"
stratalabs	string corresponding to the labels of the covariate, when NULL will use the levels of the covariate
strataname	String of the covariate name default is nicename(cov)
stratalabs.table	String corresponding to the levels of the covariate for the number at risk table, when NULL will use the levels of the covariate. Can use a string of "-" when the labels are long
strataname.table	String of the covariate name for the number at risk table default is nicename(cov)
median.text	boolean to specify if you want the median values added to the legend (or as added text if there are no covariates), for KM only
median.lines	boolean to specify if you want the median values added as lines to the plot, for KM only



median.CI	boolean to specify if you want the 95\ with the median text (Only for KM)
set.time.text	string for the text to add survival at a specified time (eg. year OS)
set.time.line	boolean to specify if you want the survival added as lines to the plot at a specified point
set.time	Numeric values of the specific time of interest, default is 5 (Multiple values can be entered)
set.time.CI	boolean to specify if you want the 95\ interval with the set time text
censor.marks	logical value. If TRUE, includes censor marks (only for KM curves)
censor.size	size of censor marks, default is 3
censor.stroke	stroke of censor marks, default is 1.5
fsize	font size
nsize	font size for numbers in the numbers at risk table
lsize	line size
psize	size of the pvalue
median.size	size of the median text (Only when there are no covariates)
median.pos	vector of length 2 corresponding to the median position (Only when there are no covariates)
median.lsize	line size of the median lines
set.size	size of the survival at a set time text (Only when there are no covariates)
set.pos	vector of length 2 corresponding to the survival at a set point position (Only when there are no covariates)
set.lsize	line size of the survival at set points
ylim	vector of length 2 corresponding to limits of y-axis. Default to NULL
linetype	vector of line types; default is solid for all lines
xlim	vector of length 2 corresponding to limits of x-axis. Default to NULL
legend.pos	Can be either a string corresponding to the legend position ("left", "top", "right", "bottom", "none") or a vector of length 2 corresponding to the legend position (uses normalized units (ie the c(0.5,0.5) is the middle of the plot))
pval.pos	vector of length 2 corresponding to the p-value position
event	String specifying if the event should be mapped to the colour, or linetype when plotting both events to colour = "col", line type
flip.CIF	boolean to flip the CIF curve to start at 1
cut	numeric value indicating where to divide a continuous covariate (default is the median)
eventlabs	String corresponding to the event type names
event.name	String corresponding to the label of the event types
Numbers_at_risk_text	String for the label of the number at risk
HR.digits	Number of digits printed of the hazard ratio

HR.pval.digits Number of digits printed of the hazard ratio pvalue  
 pval.digits Number of digits printed of the Gray's/log rank pvalue  
 median.digits Number of digits printed of the median pvalue  
 set.time.digits  
 Number of digits printed of the probability at a specified time  
 print.n.missing  
 Logical, should the number of missing be shown !Needs to be checked

---

 ggkmcif\_paste

*Plot KM and CIF curves with ggplot*


---

## Description

This function puts together a survival curve, and a number at risk table

## Usage

```
ggkmcif_paste(list_gg)
```

## Arguments

list\_gg list containing the results of ggkmcif

## Value

a gtable with three elements, the survival curve, a spacer and the number at risk table

## Examples

```

data("pembrolizumab")
plot <- ggkmcif(response=c('pfs_time', 'pfs_status'),
data=pembrolizumab, returns = TRUE)

# Highlighting a section of the curve
plot[[1]] <- plot[[1]] +
ggplot2::geom_rect(xmin=4, xmax=8, ymin=0.15, ymax=0.4, alpha=0.01, fill='yellow')

# Putting the curve back together
ggkmcif_paste(plot)

```

---

hbold	<i>Bold strings in HTML</i>
-------	-----------------------------

---

**Description**

Bold strings in HTML

**Usage**

```
hbold(strings)
```

**Arguments**

strings	A vector of strings to bold.
---------	------------------------------

---

---

lbold	<i>Bold strings in LaTeX</i>
-------	------------------------------

---

**Description**

Bold strings in LaTeX

**Usage**

```
lbold(strings)
```

**Arguments**

strings	A vector of strings to bold.
---------	------------------------------

---

---

lpvalue	<i>Formats p-values for LaTeX</i>
---------	-----------------------------------

---

**Description**

Returns <0.001 if pvalue is <0.001. Else rounds the pvalue to specified significant digits. Will bold the p-value if it is <= 0.05

**Usage**

```
lpvalue(x, sigdigits = 2)
```

**Arguments**

x	an integer
sigdigits	number of significant digit to report

mvsum

*Get multivariate summary dataframe***Description**

Returns a dataframe with the model summary and global p-value for multi-level variables.

**Usage**

```
mvsum(
  model,
  data,
  digits = getOption("reportRmd.digits", 2),
  showN = TRUE,
  showEvent = TRUE,
  markup = TRUE,
  sanitize = TRUE,
  nicenames = TRUE,
  CIwidth = 0.95,
  vif = TRUE
)
```

**Arguments**

model	fitted model object
data	dataframe containing data
digits	number of digits to round to
showN	boolean indicating sample sizes should be shown for each comparison, can be useful for interactions
showEvent	boolean indicating if number of events should be shown. Only available for logistic.
markup	boolean indicating if you want latex markup
sanitize	boolean indicating if you want to sanitize all strings to not break LaTeX
nicenames	boolean indicating if you want to replace . and _ in strings with a space.
CIwidth	width for confidence intervals, defaults to 0.95
vif	boolean indicating if the variance inflation factor should be included. See details

**Details**

Global p-values are likelihood ratio tests for lm, glm and polr models. For lme models an attempt is made to re-fit the model using ML and if successful LRT is used to obtain a global p-value. For coxph models the model is re-run without robust variances with and without each variable and a LRT is presented. If unsuccessful a Wald p-value is returned. For GEE and CRR models Wald global p-values are returned.

If the variance inflation factor is requested (VIF=T) then a generalised VIF will be calculated in the same manner as the car package.

VIF for competing risk models is computed by fitting a linear model with a dependent variable comprised of the sum of the model independent variables and then calculating VIF from this linear model.

## References

John Fox & Georges Monette (1992) Generalized Collinearity Diagnostics, Journal of the American Statistical Association, 87:417, 178-183, DOI: 10.1080/01621459.1992.10475190

John Fox and Sanford Weisberg (2019). An R Companion to Applied Regression, Third Edition. Thousand Oaks CA: Sage.

---

nestTable	<i>Combine two table columns into a single column with levels of one nested within levels of the other.</i>
-----------	---

---

## Description

This function accepts a data frame (via the data argument) and combines two columns into a single column with values from the head\_col serving as headers and values of the to\_col displayed underneath each header. The resulting table is then passed to outTable for printing and output, to use the grouped table as a data frame specify tableOnly=TRUE. By default the headers will be bolded and the remaining values indented.

## Usage

```
nestTable(
  data,
  head_col,
  to_col,
  colHeader = "",
  caption = NULL,
  indent = TRUE,
  boldheaders = TRUE,
  hdr_prefix = "",
  hdr_suffix = "",
  digits = getOption("reportRmd.digits", 2),
  tableOnly = FALSE,
  fontsize
)
```

## Arguments

data	dataframe
head_col	character value specifying the column name with the headers

to_col	character value specifying the column name to add the headers into
colHeader	character with the desired name of the first column. The default is to leave this empty for output or, for table only output to use the column name 'col1'.
caption	table caption
indent	Boolean should the original values in the to_col be indented
boldheaders	Boolean should the header column values be bolded
hdr_prefix	character value that will prefix headers
hdr_suffix	character value that will suffix headers
digits	number of digits to round numeric columns to, wither a single number or a vector corresponding to the number of numeric columns
tableOnly	boolean indicating if the table should be formatted for printing or returned as a data frame
fontsize	PDF/HTML output only, manually set the table fontsize

### Details

Note that it is possible to combine multiple tables (more than two) with this function.

### Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

### Examples

```
## Investigate models to predict baseline ctDNA and tumour size and display together
## (not clinically useful!)
data(pembrolizumab)
fit1 <- lm(baseline_ctdna~age+l_size+pd11,data=pembrolizumab)
m1 <- rm_mvsum(fit1,tableOnly=TRUE)
m1$Response = 'ctDNA'
fit2 <- lm(l_size~age+baseline_ctdna+pd11,data=pembrolizumab)
m2 <- rm_mvsum(fit2,tableOnly=TRUE)
m2$Response = 'Tumour Size'
nestTable(rbind(m1,m2),head_col='Response',to_col='Covariate')
```

---

nicename

*Lean strings for printing*

---

### Description

Returns strings with . and \_ replaced by a space. This is nice when printing column names of your dataframe in a report

**Usage**

```
nicename(strings, check_numbers = TRUE)
```

**Arguments**

strings            vector of strings to give a nice name  
 check\_numbers    boolean indicating if numbers with decimals should be checked for and retained.

---

niceNum            *Round retaining digits*

---

**Description**

Round retaining digits

**Usage**

```
niceNum(x, digits = 2)
```

**Arguments**

x                    a vector  
 digits              numeric

---

outTable            *Print tables to PDF/Latex HTML or Word*

---

**Description**

Output the table nicely to whatever format is appropriate. This is the output function used by the `rm_*` printing functions.

**Usage**

```
outTable(
  tab,
  row.names = NULL,
  to_indent = numeric(0),
  bold_headers = TRUE,
  rows_bold = numeric(0),
  bold_cells = NULL,
  caption = NULL,
  digits = getOption("reportRmd.digits", 2),
  align,
  applyAttributes = TRUE,
```

```

    keep.rownames = FALSE,
    nicenames = TRUE,
    fontsize,
    chunk_label,
    format = NULL
  )

```

### Arguments

tab	a table to format
row.names	a string specifying the column name to assign to the rownames. If NULL (the default) then rownames are removed.
to_indent	numeric vector indicating which rows to indent in the first column.
bold_headers	boolean indicating if the column headers should be bolded
rows_bold	numeric vector indicating which rows to bold
bold_cells	array indices indicating which cells to bold. These will be in addition to rows bolded by rows_bold.
caption	table caption
digits	number of digits to round numeric columns to, wither a single number or a vector corresponding to the number of numeric columns in tab
align	string specifying column alignment, defaults to left alignment of the first column and right alignment of all other columns. The align argument accepts a single string with 'l' for left, 'c' for centre and 'r' for right, with no separations. For example, to set the left column to be centred, the middle column right-aligned and the right column left aligned use: align='crl'
applyAttributes	boolean indicating if the function should use to_indent and bold_cells formatting attributes. This will only work properly if the dimensions of the table output from rm_covsum, rm_uvsum etc haven't changed.
keep.rownames	should the row names be included in the output
nicenames	boolean indicating if you want to replace . and _ in strings with a space
fontsize	PDF/HTML output only, manually set the table fontsize
chunk_label	only used knitting to Word docs to allow cross-referencing
format	if specified ('html','latex') will override the global pandoc setting

### Details

Entire rows can be bolded, or specific cells. Currently indentation refers to the first column only. By default, underscores in column names are converted to spaces. To disable this set rm\_ to FALSE

### Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned



**Examples**

```
# To make custom changes or change the fontsize in PDF/HTML
data("pembrolizumab")
tab <- rm_covsum(data=pembrolizumab,maincov = 'change_ctdna_group',
covs=c('age', 'sex', 'pd11', 'tmb', 'l_size'),show.tests=TRUE,tableOnly = TRUE)
outTable(tab, fontsize=7)

# To bold columns with the variable names
rows_bold <- c(1,4,7,10,13)
outTable(tab,rows_bold = rows_bold)

# To bold the estimates for male/female
bold_cells <- as.matrix(expand.grid(5:6,1:ncol(tab)))
outTable(tab,bold_cells= bold_cells)

# Output the above table to HTML or LaTeX
#cat(outTable(tab=tab)) #Knits to specified global setting
#cat(outTable(tab, format="html"), file = "tab.html") #HTML output
#cat(outTable(tab, format="latex"), file = "tab.tex") #LaTeX output
```

---

pembrolizumab	<i>Survival data Survival status and ctDNA levels for patients receiving pembrolizumab</i>
---------------	--

---

**Description**

Survival data

Survival status and ctDNA levels for patients receiving pembrolizumab

**Usage**

```
pembrolizumab
```

**Format**

A data frame with 94 rows and 15 variables:

**id** Patient ID

**age** Age at study entry

**sex** Patient Sex

**cohort** Study Cohort: A = Squamous cell carcinoma of soft pallate, B = Triple negative breast cancer, C = Ovarian, high grade serous, D = Melanoma, E = Other Solid Tumor

**l\_size** Target lesion size at baseline

**pd11** PD L1 percent

**tmb** log of TMB

**baseline\_ctdna** Baseline ctDNA

**change\_ctdna\_group** Did ctDNA increase or decrease from baseline to cycle 3

**orr** Objective Response

**cbr** Clinical Beneficial Response

**os\_status** Overall survival status, 0 = alive, 1 = deceased

**os\_time** Overall survival time in months

**pfs\_status** Progression free survival status, 0 = progression free, 1 = progressed

**pfs\_time** Progression free survival time in months

### Source

<https://www.nature.com/articles/s43018-020-0096-5>

---

plotuv

*Plot multiple bivariate relationships in a single plot*

---

### Description

This function is designed to accompany `uvsum` as a means of visualising the results, and uses similar syntax.

### Usage

```
plotuv(
  response,
  covs,
  data,
  showN = FALSE,
  showPoints = TRUE,
  na.rm = TRUE,
  response_title = NULL,
  return_plotlist = FALSE,
  ncol = 2,
  p_margins = c(0, 0.2, 1, 0.2),
  bpThreshold = 20,
  mixed = TRUE
)
```

### Arguments

<code>response</code>	character vector with names of columns to use for response
<code>covs</code>	character vector with names of columns to use for covariates
<code>data</code>	dataframe containing your data
<code>showN</code>	boolean indicating whether sample sizes should be shown on the plots
<code>showPoints</code>	boolean indicating whether individual data points should be shown when $n > 20$ in a category

<code>na.rm</code>	boolean indicating whether na values should be shown or removed
<code>response_title</code>	character value with title of the plot
<code>return_plotlist</code>	boolean indicating that the list of plots should be returned instead of a plot, useful for applying changes to the plot, see details
<code>ncol</code>	the number of columns of plots to be display in the <code>ggarrange</code> call, defaults to 2
<code>p_margins</code>	sets the TRBL margins of the individual plots, defaults to <code>c(0,0.2,1,.2)</code>
<code>bpThreshold</code>	Default is 20, if there are fewer than 20 observations in a category then dotplots, as opposed to boxplots are shown.
<code>mixed</code>	should a mix of dotplots and boxplots be shown based on sample size? If false then all categories will be shown as either dotplots, or boxplots according the <code>bpThreshold</code> and the smallest category size

### Details

Plots are displayed as follows: If response is continuous For a numeric predictor scatterplot For a categorical predictor: If 20+ observations available boxplot, otherwise dotplot with median line If response is a factor For a numeric predictor: If 20+ observations available boxplot, otherwise dotplot with median line For a categorical predictor barplot Response variables are shown on the ordinate (y-axis) and covariates on the abscissa (x-axis)

### Value

a list containing plots for each variable in `covs`

a plot object

### See Also

[ggplot](#) and [ggarrange](#)

### Examples

```
## Run multiple univariate analyses on the pembrolizumab dataset to predict cbr and
## then visualise the relationships.
data("pembrolizumab")
rm_uvsum(data=pembrolizumab,
response='cbr',covs=c('age','sex','l_size','baseline_ctdna'))
plotuv(data=pembrolizumab, response='cbr',
covs=c('age','sex','l_size','baseline_ctdna'),showN=TRUE)
```

---

psthr *Round and paste with parentheses*

---

**Description**

Round and paste with parentheses

**Usage**

```
psthr(x, y = 2)
```

**Arguments**

x                    a numeric vector  
y                    integer corresponding to the number of digits to round by

---

pstprn *Paste with parentheses*

---

**Description**

Paste with parentheses

**Usage**

```
pstprn(x)
```

**Arguments**

x                    a vector

---

pvalue *Formats p-values*

---

**Description**

Returns <0.001 if pvalue is <0.001. Else rounds the pvalue to specified significant digits

**Usage**

```
pvalue(x, digits)
```

**Arguments**

x                    an integer  
digits                the number of significant digits to return

---

rmds	<i>Replace dollar signs with html for proper HTML output</i>
------	--

---

**Description**

Replace dollar signs with html for proper HTML output

**Usage**

```
rmds(s)
```

**Arguments**

s                    a character vector

---

rm_cifsum	<i>Summarize cumulative incidence by group</i>
-----------	--

---

**Description**

Displays event counts and event rates at specified time points for the entire cohort and by group. Gray's test of differences in cumulative incidence is displayed.

**Usage**

```
rm_cifsum(
  data,
  time,
  status,
  group = NULL,
  eventcode = 1,
  cencode = 0,
  eventtimes,
  eventtimeunit,
  eventtimeLbls = NULL,
  CIwidth = 0.95,
  unformattedp = FALSE,
  na.action = "na.omit",
  showCounts = TRUE,
  showGraystest = TRUE,
  digits = 2,
  caption = NULL,
  tableOnly = FALSE
)
```

**Arguments**

data	data frame containing survival data
time	string indicating survival time variable
status	string indicating event status variable; must have at least 3 levels, e.g. 0 = censor, 1 = event, 2 = competing risk
group	string or character vector indicating the variable to group observations by
eventcode	numerical variable indicating event, default is 1
cencode	numerical variable indicating censored observation, default is 0
eventtimes	numeric vector specifying when event probabilities should be calculated
eventtimeunit	unit of time to suffix to the time column label if event probabilities are requested, should be plural
eventtimeLbels	if supplied, a vector the same length as eventtimes with descriptions (useful for displaying years with data provided in months)
CIwidth	width of the event probabilities, default is 95%
unformattedp	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Should be used in conjunction with the digits argument.
na.action	default is to omit missing values, but can be set to throw an error using na.action='na.fail'
showCounts	boolean indicating if the at risk, events and censored columns should be output, default is TRUE
showGraystest	boolean indicating Gray's test should be included in the final table, default is TRUE
digits	the number of digits to report in the event probabilities, default is 2.
caption	table caption for markdown output
tableOnly	should a dataframe or a formatted object be returned

**Value**

A character vector of the event table source code, unless tableOnly=TRUE in which case a data frame is returned

**Examples**

```
library(survival)
data(pbc)

# Event probabilities at various time points with replacement time labels
rm_cifsum(data=dbc,time='time',status='status',
eventtimes=c(1825,3650),eventtimeLbels=c(5,10),eventtimeunit='yr')

# Event probabilities by one group
rm_cifsum(data=dbc,time='time',status='status',group='trt',
eventtimes=c(1825,3650),eventtimeunit='day')
```

```
# Event probabilities by multiple groups
rm_cifsum(data=pbcc,time='time',status='status',group=c('trt','sex'),
eventtimes=c(1825,3650),eventtimeunit='day')
```

---

rm\_covsum

*Outputs a descriptive covariate table*


---

### Description

Returns a data frame corresponding to a descriptive table.

### Usage

```
rm_covsum(
  data,
  covs,
  maincov = NULL,
  caption = NULL,
  tableOnly = FALSE,
  covTitle = "",
  digits = 1,
  digits.cat = 0,
  nicenames = TRUE,
  IQR = FALSE,
  all.stats = FALSE,
  pvalue = TRUE,
  effSize = FALSE,
  p.adjust = "none",
  unformattedp = FALSE,
  show.tests = FALSE,
  testcont = c("rank-sum test", "ANOVA"),
  testcat = c("Chi-squared", "Fisher"),
  full = TRUE,
  include_missing = FALSE,
  percentage = c("column", "row"),
  dropLevels = TRUE,
  excludeLevels = NULL,
  numobs = NULL,
  fontsize,
  chunk_label
)
```

### Arguments

data                    dataframe containing data

covs	character vector with the names of columns to include in table
maincov	covariate to stratify table by
caption	character containing table caption (default is no caption)
tableOnly	Logical, if TRUE then a dataframe is returned, otherwise a formatted printed object is returned (default).
covTitle	character with the names of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'.
digits	number of digits for summarizing mean data
digits.cat	number of digits for the proportions when summarizing categorical data (default: 0)
nicenames	boolean indicating if you want to replace . and _ in strings with a space
IQR	boolean indicating if you want to display the inter quantile range (Q1,Q3) as opposed to (min,max) in the summary for continuous variables
all.stats	boolean indicating if all summary statistics (Q1,Q3 + min,max on a separate line) should be displayed. Overrides IQR.
pvalue	boolean indicating if you want p-values included in the table
effSize	boolean indicating if you want effect sizes included in the table. Can only be obtained if pvalue is also requested. Effect sizes calculated include Cramer's V for categorical variables, Cohen's d, Wilcoxon r, or Eta-squared for numeric/continuous variables.
p.adjust	p-adjustments to be performed
unformattedp	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Best used with tableOnly = T and outTable function. See examples.
show.tests	boolean indicating if the type of statistical test and effect size used should be shown in a column beside the pvalues. Ignored if pvalue=FALSE.
testcont	test of choice for continuous variables,one of <i>rank-sum</i> (default) or <i>ANOVA</i>
testcat	test of choice for categorical variables,one of <i>Chi-squared</i> (default) or <i>Fisher</i>
full	boolean indicating if you want the full sample included in the table, ignored if maincov is NULL
include_missing	Option to include NA values of maincov. NAs will not be included in statistical tests
percentage	choice of how percentages are presented, one of <i>column</i> (default) or <i>row</i>
dropLevels	logical, indicating if empty factor levels be dropped from the output, default is TRUE.
excludeLevels	a named list of covariate levels to exclude from statistical tests in the form list(varname =c('level1','level2')). These levels will be excluded from association tests, but not the table. This can be useful for levels where there is a logical skip (ie not missing, but not presented). Ignored if pvalue=FALSE.
numobs	named list overriding the number of people you expect to have the covariate
fontsize	PDF/HTML output only, manually set the table fontsize
chunk_label	only used if output is to Word to allow cross-referencing



## Details

Comparisons for categorical variables default to chi-square tests, but if there are counts of <5 then the Fisher Exact test will be used and if this is unsuccessful then a second attempt will be made computing p-values using MC simulation. If `testcont='ANOVA'` then the t-test with unequal variance will be used for two groups and an ANOVA will be used for three or more. The statistical test used can be displayed by specifying `show.tests=TRUE`.

Effect size can be obtained when p-value is requested.

Further formatting options are available using `tableOnly=TRUE` and outputting the table with a call to `outTable`.

## Value

A character vector of the table source code, unless `tableOnly=TRUE` in which case a data frame is returned

## References

Ellis, P.D. (2010) The essential guide to effect sizes: statistical power, meta-analysis, and the interpretation of research results. Cambridge: Cambridge University Press.[doi:10.1017/CBO9780511761676](https://doi.org/10.1017/CBO9780511761676)

Lakens, D. (2013) Calculating and reporting effect sizes to facilitate cumulative science: a practical primer for t-tests and ANOVAs. *Frontiers in Psychology*, 4; 863:1-12. [doi:10.3389/fpsyg.2013.00863](https://doi.org/10.3389/fpsyg.2013.00863)

## See Also

[covsum](#), [fisher.test](#), [chisq.test](#), [wilcox.test](#), [kruskal.test](#), [anova](#), [cramer\\_v](#), [eta\\_squared](#), and [outTable](#)

## Examples

```
data("pembrolizumab")
rm_covsum(data=pembrolizumab, maincov = 'orr',
covs=c('age', 'sex', 'pd11', 'tmb', 'l_size', 'change_ctdna_group'),
show.tests=TRUE)

# To Show Effect Sizes
rm_covsum(data=pembrolizumab, maincov = 'orr',
covs=c('age', 'sex'),
effSize=TRUE)

# To make custom changes or change the fontsize in PDF/HTML
tab <- rm_covsum(data=pembrolizumab,maincov = 'change_ctdna_group',
covs=c('age', 'sex', 'pd11', 'tmb', 'l_size'),show.tests=TRUE,tableOnly = TRUE)
outTable(tab, fontsize=7)

# To return unformatted p-values
tab <- rm_covsum(data=pembrolizumab, maincov = 'orr',
covs=c('age', 'sex', 'pd11', 'tmb', 'l_size', 'change_ctdna_group'),
show.tests=TRUE,unformattedp=TRUE,tableOnly=TRUE)
outTable(tab,digits=5)
outTable(tab,digits=5, applyAttributes=FALSE) # remove bold/indent
```

---

 rm\_mvsum

*Format a regression model nicely for 'Rmarkdown'*


---

### Description

Multivariable (or univariate) regression models are re-formatted for reporting and a global p-value is added for the evaluation of factor variables.

### Usage

```
rm_mvsum(
  model,
  data,
  digits = getOption("reportRmd.digits", 2),
  covTitle = "",
  showN = TRUE,
  showEvent = TRUE,
  CIwidth = 0.95,
  vif = TRUE,
  caption = NULL,
  tableOnly = FALSE,
  p.adjust = "none",
  unformattedp = FALSE,
  nicensames = TRUE,
  chunk_label,
  fontsize
)
```

### Arguments

model	model fit
data	data that model was fit on (an attempt will be made to extract this from the model)
digits	number of digits to round estimates to, does not affect p-values
covTitle	character with the names of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'.
showN	boolean indicating sample sizes should be shown for each comparison, can be useful for interactions
showEvent	boolean indicating if number of events should be shown. Only available for logistic.
CIwidth	width for confidence intervals, defaults to 0.95
vif	boolean indicating if the variance inflation factor should be included. See details
caption	table caption

tableOnly	boolean indicating if unformatted table should be returned
p.adjust	p-adjustments to be performed (Global p-values only)
unformattedp	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Should be used in conjunction with the digits argument.
nicensames	boolean indicating if you want to replace . and _ in strings with a space
chunk_label	only used if output is to Word to allow cross-referencing
fontsize	PDF/HTML output only, manually set the table fontsize

## Details

Global p-values are likelihood ratio tests for lm, glm and polr models. For lme models an attempt is made to re-fit the model using ML and if successful LRT is used to obtain a global p-value. For coxph models the model is re-run without robust variances with and without each variable and a LRT is presented. If unsuccessful a Wald p-value is returned. For GEE and CRR models Wald global p-values are returned. For negative binomial models a deviance test is used.

If the variance inflation factor is requested (VIF=T) then a generalised VIF will be calculated in the same manner as the car package.

The number of decimal places to display the statistics can be changed with digits, but this will not change the display of p-values. If more significant digits are required for p-values then use tableOnly=TRUE and format as desired.

## Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

## References

John Fox & Georges Monette (1992) Generalized Collinearity Diagnostics, Journal of the American Statistical Association, 87:417, 178-183, doi:10.1080/01621459.1992.10475190

John Fox and Sanford Weisberg (2019). An R Companion to Applied Regression, Third Edition. Thousand Oaks CA: Sage.

## Examples

```
data("pembrolizumab")
glm_fit = glm(change_ctdna_group~sex:age+baseline_ctdna+l_size,
data=pembrolizumab,family = 'binomial')
rm_mvsum(glm_fit)

#linear model with p-value adjustment
lm_fit=lm(baseline_ctdna~age+sex+l_size+tmb,data=pembrolizumab)
rm_mvsum(lm_fit,p.adjust = "bonferroni")
#Coxph
require(survival)
res.cox <- coxph(Surv(os_time, os_status) ~ sex+age+l_size+tmb, data = pembrolizumab)
rm_mvsum(res.cox, vif=TRUE)
```

---

rm_survdiff	<i>Display event counts, expected event counts and logrank test of differences</i>
-------------	--

---

### Description

This is a wrapper function around the `survdiff` function to display overall event rates and group-specific rates along with the log-rank test of a difference in survival between groups in a single table suitable for markdown output. Median survival times are included by default but can be removed setting `median=FALSE`

### Usage

```
rm_survdiff(
  data,
  time,
  status,
  covs,
  strata,
  includeVarNames = FALSE,
  digits = 1,
  showCols = c("N", "Observed", "Expected"),
  CIwidth = 0.95,
  conf.type = "log",
  caption = NULL,
  tableOnly = FALSE,
  fontsize
)
```

### Arguments

<code>data</code>	data frame containing survival data
<code>time</code>	string indicating survival time variable
<code>status</code>	string indicating event status variable
<code>covs</code>	character vector indicating variables to group observations by
<code>strata</code>	string indicating the variable to stratify observations by
<code>includeVarNames</code>	boolean indicating if the variable names should be included in the output table, default is <code>FALSE</code>
<code>digits</code>	the number of digits in the survival rate
<code>showCols</code>	character vector indicating which of the optional columns to display, defaults to <code>c("N", "Observed", "Expected")</code>
<code>CIwidth</code>	width of the median survival estimates, default is <code>95%</code>
<code>conf.type</code>	type of confidence interval see <a href="#">survfit</a> for details. Default is <code>'log'</code> .

caption	table caption
tableOnly	should a dataframe or a formatted object be returned
fontsize	PDF/HTML output only, manually set the table fontsize

**Value**

A character vector of the survival table source code, unless tableOnly=TRUE in which case a data frame is returned

**See Also**

[survdiff](#)

**Examples**

```
#' # Differences between sex
data("pembrolizumab")
rm_survdiff(data=pembrolizumab,time='os_time',status='os_status',
covs='sex',digits=1)

# Differences between sex, stratified by cohort
rm_survdiff(data=pembrolizumab,time='os_time',status='os_status',
covs='sex',strata='cohort',digits=1)
# Differences between sex/cohort groups
rm_survdiff(data=pembrolizumab,time='os_time',status='os_status',
covs=c('sex','cohort'),digits=1)
```

---

rm\_survsum

*Summarise survival data by group*

---

**Description**

Displays event counts, median survival time and survival rates at specified times points for the entire cohort and by group. The logrank test of differences in survival curves is displayed.

**Usage**

```
rm_survsum(
  data,
  time,
  status,
  group = NULL,
  survtimes = NULL,
  survtimeunit,
  survtimesLb1s = NULL,
  CIwidth = 0.95,
  unformattedp = FALSE,
  conf.type = "log",
```

```

na.action = "na.omit",
showCounts = TRUE,
showLogrank = TRUE,
eventProb = FALSE,
digits = getOption("reportRmd.digits", 2),
caption = NULL,
tableOnly = FALSE,
fontsize
)

```

### Arguments

<code>data</code>	data frame containing survival data
<code>time</code>	string indicating survival time variable
<code>status</code>	string indicating event status variable
<code>group</code>	string or character vector indicating the variable(s) to group observations by. If this is left as NULL (the default) then summaries are provided for the entire cohort.
<code>survtimes</code>	numeric vector specifying when survival probabilities should be calculated.
<code>survtimeunit</code>	unit of time to suffix to the time column label if survival probabilities are requested, should be plural
<code>survtimesLbels</code>	if supplied, a vector the same length as <code>survtimes</code> with descriptions (useful for displaying years with data provided in months)
<code>CIwidth</code>	width of the survival probabilities, default is 95%
<code>unformattedp</code>	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Should be used in conjunction with the <code>digits</code> argument.
<code>conf.type</code>	type of confidence interval see <a href="#">survfit</a> for details. Default is 'log'.
<code>na.action</code>	default is to omit missing values, but can be set to throw an error using <code>na.action='na.fail'</code>
<code>showCounts</code>	boolean indicating if the at risk, events and censored columns should be output; default is TRUE
<code>showLogrank</code>	boolean indicating if the log-rank test statistic and p-value should be output; default is TRUE
<code>eventProb</code>	boolean indicating if event probabilities, rather than survival probabilities, should be displayed; default is FALSE
<code>digits</code>	the number of digits in the survival rate, default is 2, unless the <code>reportRmd.digits</code> option is set
<code>caption</code>	table caption for markdown output
<code>tableOnly</code>	should a dataframe or a formatted object be returned
<code>fontsize</code>	PDF/HTML output only, manually set the table fontsize

### Details

This summary table is supplied for simple group comparisons only. To examine differences in groups with stratification see [rm\\_survdiff](#). To summarise differences in survival rates controlling for covariates see [rm\\_survtime](#).

**Value**

A character vector of the survival table source code, unless tableOnly=TRUE in which case a data frame is returned

**See Also**

[survfit](#)

**Examples**

```
# Simple median survival table
data("pembrolizumab")
rm_survsum(data=pembrolizumab,time='os_time',status='os_status')

# Survival table with yearly survival rates
rm_survsum(data=pembrolizumab,time='os_time',status='os_status',
survtimes=c(12,24),survtimesLbls=1:2, survtimeunit='yr')

#Median survival by group
rm_survsum(data=pembrolizumab,time='os_time',status='os_status',group='sex')

# Survival Summary by cohort, displayed in years
rm_survsum(data=pembrolizumab,time='os_time',status='os_status',
group="cohort",survtimes=seq(12,72,12),
survtimesLbls=seq(1,6,1),
survtimeunit='years')

# Survival Summary by Sex and ctDNA group
rm_survsum(data=pembrolizumab,time='os_time',status='os_status',
group=c('sex','change_ctdna_group'),survtimes=c(12,24),survtimeunit='mo')
```

---

rm\_survtime

*Display survival rates and events for specified times*

---

**Description**

This is a wrapper for the survfit function to output a tidy display for reporting. Either Kaplan Meier or Cox Proportional Hazards models may be used to estimate the survival probabilities.

**Usage**

```
rm_survtime(
  data,
  time,
  status,
  covs = NULL,
  strata = NULL,
  type = "KM",
```

```

survtimes,
survtimeunit,
strata.prefix = NULL,
survtimesLb1s = NULL,
showCols = c("At Risk", "Events", "Censored"),
CIwidth = 0.95,
conf.type = "log",
na.action = "na.omit",
showCounts = TRUE,
digits = getOption("reportRmd.digits", 2),
caption = NULL,
tableOnly = FALSE,
fontsize
)

```

### Arguments

<code>data</code>	data frame containing survival data
<code>time</code>	string indicating survival time variable
<code>status</code>	string indicating event status variable
<code>covs</code>	character vector with the names of variables to adjust for in coxph fit
<code>strata</code>	string indicating the variable to group observations by. If this is left as NULL (the default) then event counts and survival rates are provided for the entire cohort.
<code>type</code>	survival function, if no covs are specified defaults to Kaplan-Meier, otherwise the Cox PH model is fit. Use <code>type='PH'</code> to fit a Cox PH model with no covariates.
<code>survtimes</code>	numeric vector specifying when survival probabilities should be calculated.
<code>survtimeunit</code>	unit of time to suffix to the time column label if survival probabilities are requested, should be plural
<code>strata.prefix</code>	character value describing the grouping variable
<code>survtimesLb1s</code>	if supplied, a vector the same length as <code>survtimes</code> with descriptions (useful for displaying years with data provided in months)
<code>showCols</code>	character vector specifying which of the optional columns to display, defaults to <code>c('At Risk','Events','Censored')</code>
<code>CIwidth</code>	width of the survival probabilities, default is 95%
<code>conf.type</code>	type of confidence interval see <a href="#">survfit</a> for details. Default is 'log'.
<code>na.action</code>	default is to omit missing values, but can be set to throw an error using <code>na.action='na.fail'</code>
<code>showCounts</code>	boolean indicating if the at risk, events and censored columns should be output, default is TRUE
<code>digits</code>	the number of digits in the survival rate, default is 2.
<code>caption</code>	table caption for markdown output
<code>tableOnly</code>	should a dataframe or a formatted object be returned
<code>fontsize</code>	PDF/HTML output only, manually set the table fontsize



**Details**

If covariates are supplied then a Cox proportional hazards model is fit for the entire cohort and each strata. Otherwise the default is for Kaplan-Meier estimates. Setting `type = 'PH'` will force a proportional hazards model.

**Value**

A character vector of the survival table source code, unless `tableOnly=TRUE` in which case a data frame is returned

**See Also**

[survfit](#)

**Examples**

```
# Kaplan-Meier survival probabilities with time displayed in years
data("pembrolizumab")
rm_survtime(data=pembrolizumab,time='os_time',status='os_status',
strata="cohort",type='KM',survtimes=seq(12,72,12),
survtimesLbls=seq(1,6,1),
survtimeunit='years')

# Cox Proportional Hazards survival probabilities
rm_survtime(data=pembrolizumab,time='os_time',status='os_status',
strata="cohort",type='PH',survtimes=seq(12,72,12),survtimeunit='months')

# Cox Proportional Hazards survival probabilities controlling for age
rm_survtime(data=pembrolizumab,time='os_time',status='os_status',
covs='age',strata="cohort",survtimes=seq(12,72,12),survtimeunit='months')
```

---

rm\_uvsum

*Output several univariate models nicely in a single table*

---

**Description**

A table with the model parameters from running separate univariate models on each covariate. For factors with more than two levels a Global p-value is returned.

**Usage**

```
rm_uvsum(
  response,
  covs,
  data,
  digits = getOption("reportRmd.digits", 2),
  covTitle = "",
```

```

caption = NULL,
tableOnly = FALSE,
removeInf = FALSE,
p.adjust = "none",
unformattedp = FALSE,
chunk_label,
gee = FALSE,
id = NULL,
corstr = NULL,
family = NULL,
type = NULL,
offset,
strata = 1,
nicenames = TRUE,
showN = TRUE,
showEvent = TRUE,
CIwidth = 0.95,
reflevel = NULL,
returnModels = FALSE,
fontsize,
forceWald
)

```

### Arguments

response	string vector with name of response
covs	character vector with the names of columns to fit univariate models to
data	dataframe containing data
digits	number of digits to round estimates and CI to. Does not affect p-values.
covTitle	character with the names of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'.
caption	character containing table caption (default is no caption)
tableOnly	boolean indicating if unformatted table should be returned
removeInf	boolean indicating if infinite estimates should be removed from the table
p.adjust	p-adjustments to be performed (Global p-values only)
unformattedp	boolean indicating if you would like the p-value to be returned unformatted (ie not rounded or prefixed with '<'). Should be used in conjunction with the digits argument.
chunk_label	only used if output is to Word to allow cross-referencing
gee	boolean indicating if gee models should be fit to account for correlated observations. If TRUE then the id argument must specify the column in the data which indicates the correlated clusters.
id	character vector which identifies clusters. Only used for geeglm

corstr	character string specifying the correlation structure. Only used for geeglm. The following are permitted: ""independence"", ""exchangeable"", ""ar1"", ""unstructured"" and ""userdefined""
family	description of the error distribution and link function to be used in the model. Only used for geeglm
type	string indicating the type of univariate model to fit. The function will try and guess what type you want based on your response. If you want to override this you can manually specify the type. Options include "linear", "logistic", "poisson", "coxph", "crr", "boxcox", "ordinal", "geeglm"
offset	string specifying the offset term to be used for Poisson or negative binomial regression. Example: offset="log(follow_up)"
strata	character vector of covariates to stratify by. Only used for coxph and crr
nicenames	boolean indicating if you want to replace . and _ in strings with a space
showN	boolean indicating if you want to show sample sizes
showEvent	boolean indicating if you want to show number of events. Only available for logistic.
CIwidth	width of confidence interval, default is 0.95
reflevel	manual specification of the reference level. Only used for ordinal regression. This will allow you to see which model is not fitting if the function throws an error
returnModels	boolean indicating if a list of fitted models should be returned. If this is TRUE then the models will be returned, but the output will be suppressed. In addition to the model elements a data element will be appended to each model so that the fitted data can be examined, if necessary. See Details
fontsize	PDF/HTML output only, manually set the table fontsize
forceWald	boolean indicating if Wald confidence intervals should be used instead of profile likelihood. This is not recommended, but can speed up computations. To use throughout a document use options(reportRmd.forceWald=TRUE)

### Details

Global p-values are likelihood ratio tests for lm, glm and polr models. For lme models an attempt is made to re-fit the model using ML and if successful LRT is used to obtain a global p-value. For coxph models the model is re-run without robust variances with and without each variable and a LRT is presented. If unsuccessful a Wald p-value is returned. For GEE and CRR models Wald global p-values are returned.

The number of decimal places to display the statistics can be changed with digits, but this will not change the display of p-values. If more significant digits are required for p-values then use tableOnly=TRUE and format as desired.

### Value

A character vector of the table source code, unless tableOnly=TRUE in which case a data frame is returned

**See Also**

[uvsum](#), [lm](#), [glm](#), [crr](#), [coxph](#), [lme](#), [geeglm](#), [polr](#)

**Examples**

```
# Examples are for demonstration and are not meaningful
# Coxph model with 90% CI
data("pembrolizumab")
rm_uvsum(response = c('os_time', 'os_status'),
covs=c('age', 'sex', 'baseline_ctdna', 'l_size', 'change_ctdna_group'),
data=pembrolizumab, CIwidth=.9)

# Linear model with default 95% CI
rm_uvsum(response = 'baseline_ctdna',
covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
data=pembrolizumab)

# Logistic model with default 95% CI
rm_uvsum(response = 'os_status',
covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
data=pembrolizumab, family = binomial)
# Poisson models returned as model list
mList <- rm_uvsum(response = 'baseline_ctdna',
covs=c('age', 'sex', 'l_size', 'pd11', 'tmb'),
data=pembrolizumab, returnModels=TRUE)
#'
# GEE on correlated outcomes
data("ctDNA")
rm_uvsum(response = 'size_change',
covs=c('time', 'ctdna_status'),
gee=TRUE,
id='id', corstr="exchangeable",
family=gaussian("identity"),
data=ctDNA, showN=TRUE)
```

---

rm\_uv\_mv

*Combine univariate and multivariable regression tables*

---

**Description**

This function will combine `rm_uvsum` and `rm_mvsum` outputs into a single table. The `tableOnly` argument must be set to `TRUE` when tables to be combined are created. The resulting table will be in the same order as the `uvsum` table and will contain the same columns as the `uvsum` and `mvsum` tables, but the p-values will be combined into a single column. There must be a variable overlapping between the `uvsum` and `mvsum` tables and all variables in the `mvsum` table must also appear in the `uvsum` table.

**Usage**

```
rm_uv_mv(
  uvsumTable,
  mvsumTable,
  covTitle = "",
  vif = FALSE,
  showN = FALSE,
  showEvent = FALSE,
  caption = NULL,
  tableOnly = FALSE,
  chunk_label,
  fontsize
)
```

**Arguments**

uvsumTable	Output from <code>rm_uvsum</code> , with <code>tableOnly=TRUE</code>
mvsumTable	Output from <code>rm_mvsum</code> , with <code>tableOnly=TRUE</code>
covTitle	character with the names of the covariate (predictor) column. The default is to leave this empty for output or, for table only output to use the column name 'Covariate'.
vif	boolean indicating if the variance inflation factor should be shown if present in the mvsumTable. Default is FALSE.
showN	boolean indicating if sample sizes should be displayed.
showEvent	boolean indicating if number of events (dichotomous outcomes) should be displayed.
caption	table caption
tableOnly	boolean indicating if unformatted table should be returned
chunk_label	only used if output is to Word to allow cross-referencing
fontsize	PDF/HTML output only, manually set the table fontsize

**Value**

A character vector of the table source code, unless `tableOnly=TRUE` in which case a data frame is returned

**See Also**

[rm\\_uvsum](#), [rm\\_mvsum](#)

**Examples**

```
require(survival)
data("pembrolizumab")
uvTab <- rm_uvsum(response = c('os_time', 'os_status'),
  covs=c('age', 'sex', 'baseline_ctdna', 'l_size', 'change_ctdna_group'),
```

```

data=pembrolizumab,tableOnly=TRUE)
mv_surv_fit <- coxph(Surv(os_time,os_status)~age+sex+
baseline_ctdna+l_size+change_ctdna_group, data=pembrolizumab)
uvTab <- rm_mvsum(mv_surv_fit)

#linear model
uvtab<-rm_uvsum(response = 'baseline_ctdna',
covs=c('age','sex','l_size','pd11','tmb'),
data=pembrolizumab,tableOnly=TRUE)
lm_fit=lm(baseline_ctdna~age+sex+l_size+tmb,data=pembrolizumab)
mvtab<-rm_mvsum(lm_fit,tableOnly = TRUE)
rm_uv_mv(uvtab,mvtab,tableOnly=TRUE)

#logistic model
uvtab<-rm_uvsum(response = 'os_status',
covs=c('age','sex','l_size','pd11','tmb'),
data=pembrolizumab,family = binomial,tableOnly=TRUE)
logis_fit<-glm(os_status~age+sex+l_size+pd11+tmb,data = pembrolizumab,family = 'binomial')
mvtab<-rm_mvsum(logis_fit,tableOnly = TRUE)
rm_uv_mv(uvtab,mvtab,tableOnly=TRUE)

```

---

sanitizestr

*Sanitizes strings to not break LaTeX*

---

### Description

Strings with special charaters will break LaTeX if returned 'asis' by knitr. This happens every time we use one of the main reportRx functions. We first sanitize our strings with this function to stop LaTeX from breaking.

### Usage

```
sanitizestr(str)
```

### Arguments

str                    a vector of strings to sanitize

---

set\_labels

*Set variable labels*

---

### Description

Assign variable labels to a data.frame from a lookup table.

### Usage

```
set_labels(data, names_labels)
```

**Arguments**

data	data frame to be labelled
names_labels	data frame with column 1 containing variable names from data and column 2 containing variable labels. Other columns will be ignored.

**Details**

Useful if variable labels have been imported from a data dictionary. The first column in names\_labels must contain the variable name and the second column the variable label. The column names are not used.

If no label is provided then the existing label will not be changed. To remove a label set the label to NA.

**Examples**

```
# create data frame with labels
lbls <- data.frame(c1=c('cohort', 'size_change'),
                  c2=c('Cancer cohort', 'Change in tumour size'))
# set labels and return labelled data frame
set_labels(ctDNA, lbls)
```

---

set_var_labels	<i>Set variable labels</i>
----------------	----------------------------

---

**Description**

Set variable labels for a data frame using name-label pairs.

**Usage**

```
set_var_labels(data, ...)
```

**Arguments**

data	data frame containing variables to be labelled
...	Name-label pairs the name gives the name of the column in the output and the label is a character vector of length one.

**Details**

If no label is provided for a variable then the existing label will not be changed. To remove a label set the label to NA.

**Examples**

```
# set labels using name-label pairs
# and return labelled data frame
ctDNA |> set_var_labels(
  ctdna_status="detectable ctDNA",
  cohort="A cohort label")
```

---

testData	<i>Funky ctDNA data There is a weird factor with all one level, the cohort variable contains a cohort level (and Cohort A) and for one cohort all the size changes are missing</i>
----------	--

---

**Description**

Funky ctDNA data

There is a weird factor with all one level, the cohort variable contains a cohort level (and Cohort A) and for one cohort all the size changes are missing

**Usage**

```
testData
```

**Format**

A data frame with 270 rows and 6 variables:

**id** Patient ID

**cohort** Study Cohort: A = Squamous cell carcinoma of soft pallate, B = Triple negative breast cancer, C = Ovarian, high grade serous, D = Melanoma, E = Other Solid Tumor, cohort -for testing only

**badfactor** for testing

**ctdna\_status** Change in ctDNA since baseline

**time** Number of weeks on treatment

**size\_change** Percentage change in tumour measurement

**Source**

<https://www.nature.com/articles/s43018-020-0096-5>



uvsum

*Get univariate summary dataframe***Description**

Returns a dataframe corresponding to a univariate regression table

**Usage**

```
uvsum(
  response,
  covs,
  data,
  digits = getOption("reportRmd.digits", 2),
  id = NULL,
  corstr = NULL,
  family = NULL,
  type = NULL,
  offset = NULL,
  gee = FALSE,
  strata = 1,
  markup = TRUE,
  sanitize = TRUE,
  nicenames = TRUE,
  showN = TRUE,
  showEvent = TRUE,
  CIwidth = 0.95,
  refllevel = NULL,
  returnModels = FALSE,
  forceWald
)
```

**Arguments**

response	string vector with name of response
covs	character vector with the names of columns to fit univariate models to
data	dataframe containing data
digits	number of digits to round to
id	character vector which identifies clusters. Used for GEE and coxph models.
corstr	character string specifying the correlation structure. Only used for geeglm. The following are permitted: "independence", "exchangeable", "ar1", "unstructured" and "userdefined"
family	specify details of the model used. This argument does not need to be specified and should be used with caution. By default, gaussian errors are used for linear models, the binomial family with logit link is used for logistic regression and

	poisson with log link is used for poisson regression. This can be specified with the type argument, or will be inferred from the data type. See <a href="#">family</a> . Ignored for ordinal and survival regression and if the type argument is not explicitly specified.
type	string indicating the type of univariate model to fit. The function will try and guess what type you want based on your response. If you want to override this you can manually specify the type. Options include "linear", "logistic", "poisson", "coxph", "crr", "boxcox", "ordinal" and "negbin"
offset	string specifying the offset term to be used for Poisson or negative binomial regression. Example: offset="log(follow_up)"
gee	boolean indicating if gee models should be fit to account for correlated observations. If TRUE then the id argument must specify the column in the data which indicates the correlated clusters.
strata	character vector of covariates to stratify by. Only used for coxph and crr
markup	boolean indicating if you want latex markup
sanitize	boolean indicating if you want to sanitize all strings to not break LaTeX
nicenames	boolean indicating if you want to replace . and _ in strings with a space
showN	boolean indicating if you want to show sample sizes
showEvent	boolean indicating if you want to show number of events. Only available for logistic.
CIwidth	width of confidence interval, default is 0.95
reflevel	manual specification of the reference level. Only used for ordinal. This may allow you to debug if the function throws an error.
returnModels	boolean indicating if a list of fitted models should be returned.
forceWald	boolean indicating if Wald confidence intervals should be used instead of profile likelihood. This is not recommended, but can speed up computations. To use throughout a document use options(reportRmd.forceWald=TRUE)

## Details

Univariate summaries for a number of covariates, the type of model can be specified. If unspecified the function will guess the appropriate model based on the response variable.

Confidence intervals are extracted using `confint` where possible. Otherwise Student t distribution is used for linear models and the Normal distribution is used for proportions.

`returnModels` can be used to return a list of the univariate models, which will be the same length as `covs`. The data used to run each model will include all cases with observations on the response and covariate. For gee models the data are re-ordered so that the ids appear sequentially and proper estimates are given.

## See Also

[lm](#), [glm](#), [crr](#), [coxph](#), [lme](#), [geeglm](#), [polr](#), [glm.nb](#)

# Index

- \* **dataframe**
  - covsum, 5
  - mvsum, 28
  - rm\_covsum, 39
  - uvsum, 57
- \* **datasets**
  - ctDNA, 8
  - pembrolizumab, 33
  - testData, 56
- \* **helper**
  - addspace, 3
  - cap, 4
  - formatp, 16
  - hbld, 27
  - lbld, 27
  - lpvalue, 27
  - nicename, 30
  - niceNum, 31
  - psthr, 36
  - pstprn, 36
  - pvalue, 36
  - rmds, 37
  - sanitizestr, 54
- \* **model**
  - boxcoxfitRx, 3
  - crrRx, 7
- \* **plot**
  - forestplot2, 10
  - forestplotMV, 11
  - forestplotUV, 13
  - forestplotUVMV, 14
  - plotuv, 34
- addspace, 3
- anova, 7, 41
- boxcoxfitRx, 3
- cap, 4
- chisq.test, 7, 41
- clear\_labels, 4
- covsum, 5, 41
- coxph, 52, 58
- cramer\_v, 41
- crr, 7, 52, 58
- crrRx, 7
- ctDNA, 8
- eta\_squared, 41
- excelCol, 8
- excelColLetters, 9
- extract\_labels, 10
- family, 58
- fisher.test, 7, 41
- forestplot2, 10
- forestplotMV, 11
- forestplotUV, 13
- forestplotUVMV, 14
- formatp, 16
- geeglm, 52, 58
- geoR\_boxcoxfit, 16
- ggarrange, 35
- ggkmcif, 17
- ggkmcif2, 21
- ggkmcif2Parameters, 22, 23
- ggkmcif\_paste, 26
- ggplot, 35
- glm, 52, 58
- glm.nb, 58
- hbld, 27
- kruskal.test, 7, 41
- lbld, 27
- lm, 52, 58
- lme, 52, 58
- lpvalue, 27

mvsum, 28

nestTable, 29

nicename, 30

niceNum, 31

outTable, 31, 41

pembrolizumab, 33

plotuv, 34

polr, 52, 58

psthr, 36

pstprn, 36

pvalue, 36

rm\_cifsum, 37

rm\_covsum, 39

rm\_mvsum, 42, 53

rm\_survdiff, 44, 46

rm\_survsum, 45

rm\_survtime, 46, 47

rm\_uv\_mv, 52

rm\_uvsum, 49, 53

rmds, 37

sanitizestr, 54

set\_labels, 54

set\_var\_labels, 55

survdiff, 45

survfit, 44, 46–49

testData, 56

uvsum, 34, 52, 57

wilcox.test, 7, 41